Nanoemulsion coatings and use of antimicrobial ginger essential oil on citrus

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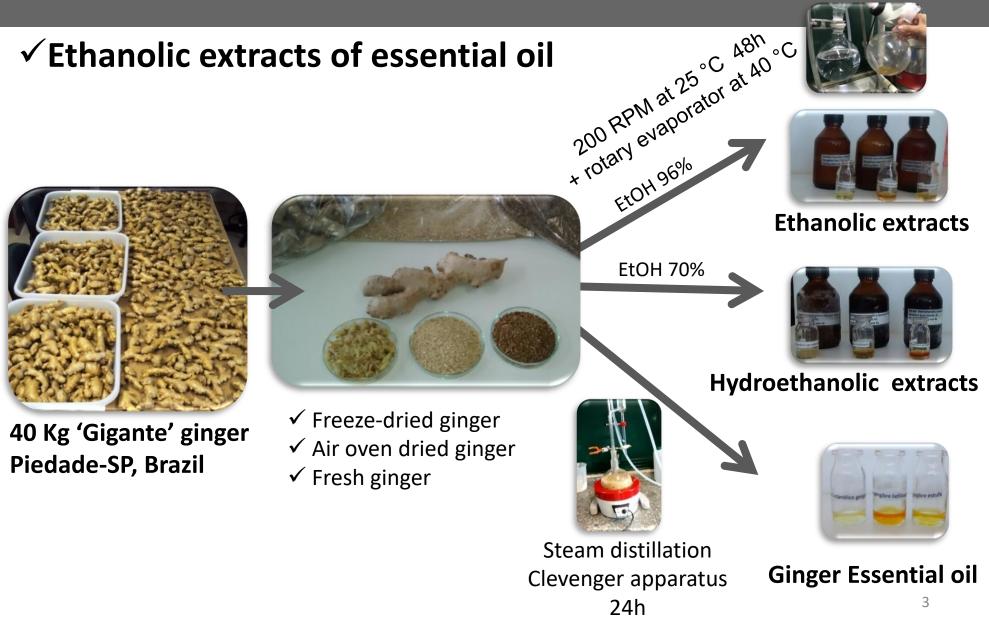




Introduction

- ✓ Shellac and carnauba wax are commonly used on citrus, carnauba is more permeable to gases, less permeable to water, shellac is more shiny but can cause off-flavor
- ✓ Carnauba wax emulsion coatings are usually microemulsions, but the smaller the lipid mycelles, the more stable and shiny the coating, thus tested a nanoemulsion carnauba coating.
- ✓ Ginger (rhizome extracts) was found to reduce microorganisms in food
- Objective was to evaluate effects of (ginger)
 antimicrobial/nanoemulsion edible coatings to preserve fruit quality and decrease decay

Antimicrobial activity: Ginger extracts



Antimicrobial activity for ginger ethanolic extracts and essential oil

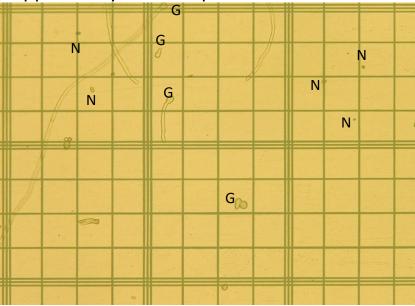
✓ Initial screening: concentration 1% and 3%

✓ Spore germination inhbition

Penicillium digitatum

•PD (potato dextrose) broth + Sample + spore suspension

Final concentration of spores solution 10⁶
Incubated at 25 °C by 120 RPM agitation
Reading after 24 hours in Neubauer chamber supported by microscope with camera



500x magnification N: not germinated; G: germinated Table 1. Percentage of germinated spores of P. digitatium,after 24 h incubation in ginger essential oil and alcoholicextracts.

		1	
	Samples	1%	3%
1	EtOH96 fresh	18.1 bc	2.9 a
2	EtOH96 freeze-dried	35.7 d	1.2 a
3	EtOH96 oven	32.2 d	0.6 a
4	EtOH70 fresh	24.6 c	25.1 b
5	EtOH70 freeze-dried	23.4 c	19.9 b
6	EtOH 70oven	14.6 b	2.3 a
7	GEO fresh	0 a	0 a
8	GEO freeze-dried	0 a	0 a
9	GEO air oven	0 a	0 a
10	GEO commercial	5.3 a	0.6 a
11	Negative control	37.4 d	37.4 c
12	Positive control	0.6 a	0.6 a
13	DMSO control	35.7 d	35.7 c
14	PVP control	36.3 d	35.7 c

Columns with different letters are significantly different by Duncan test (p<0.05) applied after Anova. Ethanolic extracts at 3% and GEO at 1 and 3% showed the highest inhibition of spore germination. Commercial GEO had lowest activity compared to extracted GEO. 4

✓ **Poisoned food technique** (test essential oils vs fungi)

Penicillium digitatum

•Samples into in solid medium (PDA) •10uL of spores suspension 10⁵

•Incubated at 25 °C

Ethanolic and Hydroethanolic extracts did not exhibit antifungal activity at 1% and 3%

GEO showed the highest mycelial zone inhibition

•Oil was dissolved in Dimethyl sulfoxide (DMSO) and alcoholic extracts in Polivinilpirrolidone (PVP)

•Tween 80 (100 μL to 100mL of medium) to disperse the sample in the medium.

Table 2. Percentage of mycelial zone inhibitioncompared to control after 5 days.

	Samples	1%	3%
1	EtOH96 fresh	*	*
2	EtOH96 freeze-dried	*	*
3	EtOH96 oven	*	*
4	EtOH70 fresh	*	*
5	EtOH70 freeze-dried	*	*
6	EtOH oven	*	*
7	GEO fresh	11.58	12.24
8	GEO freeze-dried	23.82	27.59
9	GEO air oven	10.08	21.94
10	GEO commercial	2.35	3.10
11	Negative control(water)	*	*
12	Positive control (Cyclohemide)	100.00	100.00
13	DMSO control	*	*
14	Tween 80 control	*	*
15	PVP control	*	*

*No inhibition was observed for Penicilium digitatium.

✓ Inverted Petri dish test



Figure: *Penicillium digitatum,* Neg. control: and Positive control: ammonia

•Samples placed on 6mm disc on the Petri dishes lid

•10uL of spores suspension 10⁵ on PDA medium

Incubated at 25 °C

Table 3. Percentage of mycelial zone inhibition compared to control after 5 days.

	Samples	0.5%	1%	3%
7	GEO fresh	8.8	9.4	34.1
8	GEO freeze-dried	7.9	7.3	52.6
9	GEO air oven	0	1.3	36.1
10	GEO commercial	0	5.3	11.6

Ginger oil is rich in sesquiterpenes and monoterpenes

>Zingiberene and geranial usually are the major constituents and theirs levels make the oil more potent

>Investigation into the levels of these and other compounds is underway to explain the different antifungal activity between the samples in this study

\checkmark Minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC)

- MIC Macrodilution-NCCLS (2002)
- •Samples in PD broth , spores suspension 10⁶

Ethanolic extracts: 1.0, 1.5, 2.0, 2.5, 3.0, 4.0, 5.0, 6.0 %

- •Incubated at 25 °C by 125 RPM
- •Reading after 7days





Green mold Anthracnose Penicillium digitatum Colletotrichum. gloeosporioides

0.0015%

Assessed concentrations :

GEO: 1.6, 0.8, 0.4, 0.2, 0.1, 0.5, 0.025, 0.0125, 0.0062, 0.0031, 0.0015%



1.6 0.8 0.4 0.2 0.1 0.5% Figure. MIC_GEO-fresh_*P. digitatium*

•MFC – Placed 10 uL from MIC tube and subsequent higher concentration on PDA medium ⁷

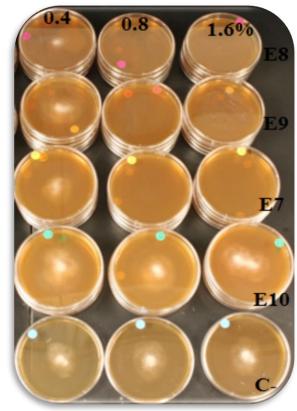


Figure. MFC_GEO- P. digitatium

Table 4. Minimum inhibitory concentration (MIC) and minimum fungicidalconcentration (MFC) of *Penicilium digitatium and Colletotrichum*gloeosporioides after 7 days of incubation.

	MIC	MFC	MIC	MFC	
Samples	Peni	icilium	Colletotrichum		
E1 Ethanolic extract-fresh ginger	2.5%	*	4.0%	6.0%	
E2 Ethanolic extract-freeze-dried	2.5%	*	2.5%	5.0%	
E3 Ethanolic extract- air oven	2.5%	*	2.5%	5.0%	
Ethanol 96% control	3.0%	*	4.0%	5.0%	
E4 HydroEthanolic extract-fresh ginger	3.0%	*	5.0%	*	
E5 HydroEthanolic extract-freeze-dried	3.0%	*	3.0%	4.0%	
E6 HydroEthanolic extract- air oven	2.5%	*	2.5%	4.0%	
Ethanol 70% control	4.0%	*	5.0%	6.0%	
E7 GEO –fresh ginger	0.4%	0.8%	0.4%	0.4%	
E8 GEO - freeze-dried ginger	0.4%	0.8%	0.1%	0.2%	
E9 GEO - air oven	0.4%	0.8%	0.8%	1.6%	
E10GEO commercial	0.8%	*	1.6%	*	

*No total inhibition at the highest concentration tested

Ethanolic and hydroethanolic extracts did not show fungicide activity against P. digitatium, however exhibited antifungal activity against C. gloeosporioides between 4 and 6%

Ethanolic and Hydroethanolic extracts did not exhibit antifungal activity in the 1% to 3% range in the poisoned food test, however they showed MIC at 2.5 and 3% in pd broth.

➢ More studies are needed to study the diffusibility and evaporation of ethanol for extracts in solid systems to improve their performance and minimize ethanol activity .

➢GEO fresh, freeze dried or air oven dried showed higher antimicrobial activity toward both microorganisms evaluated. Commercial GEO had lower activity compared to extracted GEO.

➤GEO-air oven was selected for incorporation into a nanoemulsion coating because it had the highest yield (3.5 % w/w) compared to essential oil from fresh or freeze-dried ginger (0.2 % and 3.2%, respectively).

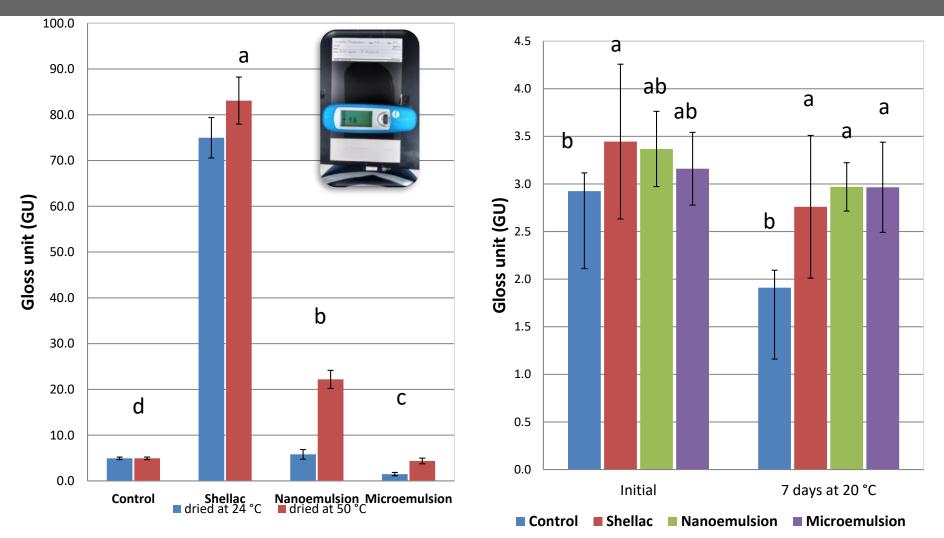
Evaluation of coatings on citrus fruit

• Experiment 1- 'Nova' mandarin

- Commercial carnauba and shellac microemulsions and an experimental carnauba nanoemulsion coating were compared to an uncoated control for fruit stored 7 d at 20 °C
- Quality analyses were done for initial (0d) and at the end of storage (7d)



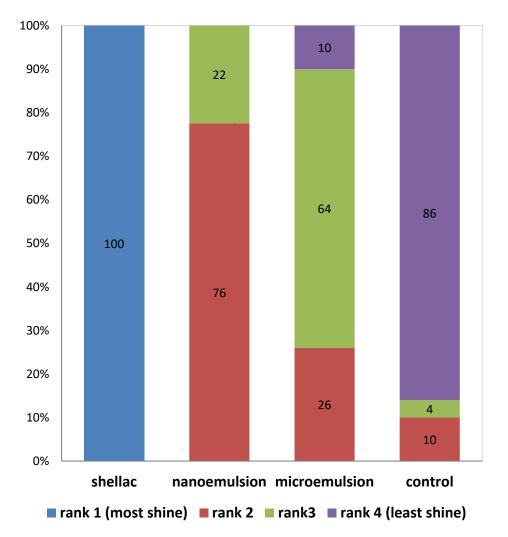
Exp.1: Quality results - gloss



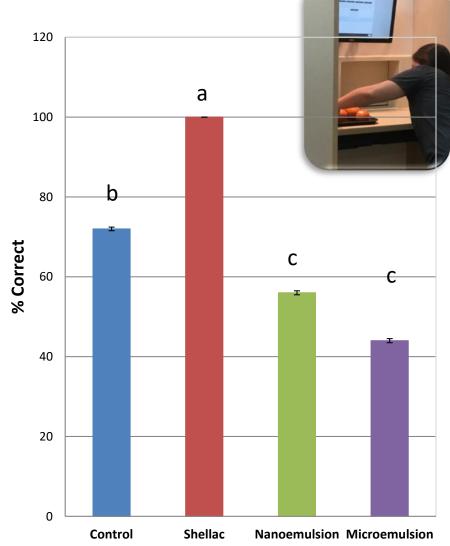
Columns with different letters are significantly different by Duncan test (p<0.05) applied after Anova.

For each storage period, columns with different letters are significantly different by Duncan test (p<0.05) applied after Anova.

Exp.1: Sensory Ranking for shine and tetrad test

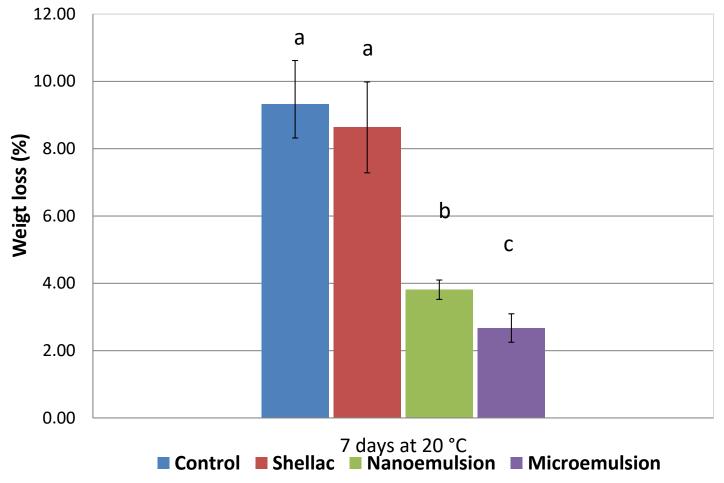


Columns with different letters are significantly different by critical absolute rank sum differences table at p<0.05 (Newell and MacFarlane, 1987),



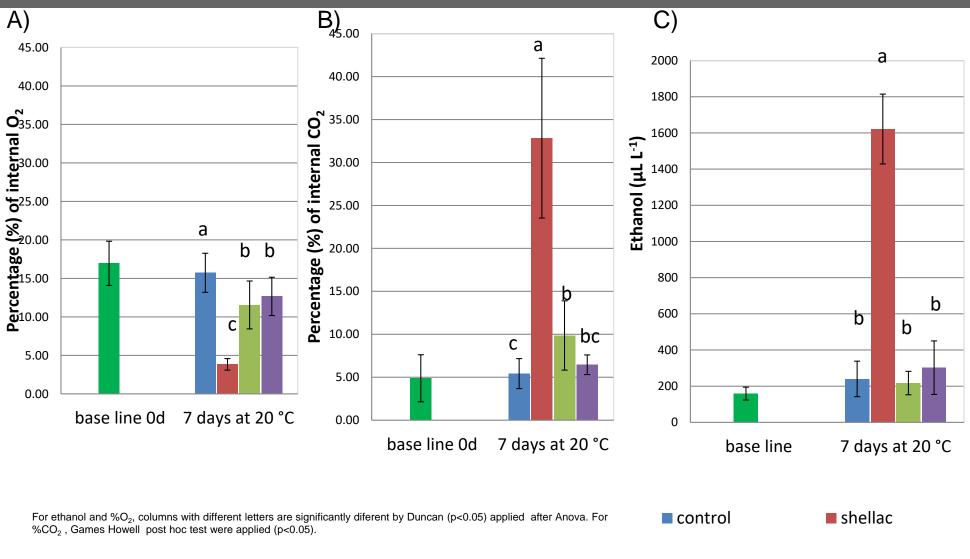
Columns with different letters are significantly different by approximation equation for tetrad (Z-test) at p=0.05,

Exp.1: Weight loss



Columns with different letters are significantly different by Games Howell (p<0.05) applied after Anova.

Exp.1: Internal CO₂, O₂ and ethanol



nanoemulsion

microemulsion

Aroma Volatiles for Nova mandarins

Class	Compound	Base line	Control	Shellac	Nano carnauba	Micro carnauba
Ciuss	compound	0 day		7 days at 20 °		cui ini uou
	Acetaldehyde	0.427	<mark>0.475* a</mark>	<mark>0.584 a</mark>	<mark>0.466 a</mark>	<mark>0.489 a</mark>
Aldahudaa	Hexanal	0.013	0.009 b <mark>c</mark>	<mark>0.007 с</mark>	<mark>0.015 a</mark>	0.013 <mark>a</mark> b
Aldehydes	Octanal	0.012	<mark>0.015 a</mark>	<mark>0.014 a</mark>	<mark>0.018 a</mark>	<mark>0.012 a</mark>
	Decanal	0.343	<mark>0.344 b</mark>	<mark>0.398 b</mark>	<mark>0.569 a</mark>	0.352 b
	Methanol	26.266	<mark>26.560 b</mark>	<mark>67.170 a</mark>	26.852 b	<mark>44.283 b</mark>
	Ethanol	158.705	240.010 b	<mark>1621.475 a</mark>	217.180 b	302.219 b
	2-MethyPropanol	0.168	<mark>0.289 b</mark>	<mark>1.753 a</mark>	<mark>0.217 b</mark>	<mark>0.350 b</mark>
	Hexanol	0.110	<mark>0.087 b</mark>	<mark>0.114 b</mark>	<mark>0.156 a</mark>	<mark>0.092 b</mark>
Alcohols	cis-3-Hexenol	0.173	<mark>0.240 b</mark>	<mark>2.671 a</mark>	<mark>0.298 b</mark>	<mark>0.323 b</mark>
AICOHOIS	trans-2-Hexenol	0.003	<mark>0.026 a</mark>	<mark>0.006 b</mark>	<mark>0.002 b</mark>	<mark>0.009 b</mark>
	Linalool	0.132	<mark>0.136 a</mark>	<mark>0.148 a</mark>	<mark>0.128 a</mark>	<mark>0.117 г</mark>
	Octanol	0.414	<mark>0.870 a</mark>	<mark>0.738 a</mark>	<mark>1.010 a</mark>	<mark>0.652 г</mark>
	Terpinen-4-ol	0.083	<mark>0.079 a</mark>	<mark>0.047 с</mark>	0.063 b	0.060 b <mark>c</mark>
	α-Terpineol	0.117	<mark>0.278 a</mark>	0.171 b	<mark>0.108 c</mark>	0.200 t
	α-Pinene	0.026	<mark>0.059 a</mark>	<mark>0.049 a</mark>	<mark>0.056 a</mark>	<mark>0.046 a</mark>
	Sabinene	0.004	<mark>0.005 b</mark>	<mark>0.005 b</mark>	<mark>0.013 a</mark>	<mark>0.005 t</mark>
Τ	Myrcene	0.170	<mark>0.297 a</mark>	<mark>0.297 a</mark>	<mark>0.358 a</mark>	0.225 a
Terpenes	Limonene	3.509	<mark>5.300 a</mark>	<mark>5.119 a</mark>	<mark>6.732 a</mark>	<mark>3.909 a</mark>
	γ-Terpinene	0.002	<mark>0.003 a</mark>	<mark>0.004 a</mark>	<mark>0.003 a</mark>	<mark>0.003 a</mark>
	Valencene	0.461	<mark>0.273 a</mark>	<mark>0.284 a</mark>	<mark>0.298 a</mark>	0.254 a
	Methyl Butanoate	0.003	<mark>0.005 b</mark>	<mark>0.038 a</mark>	<mark>0.004 b</mark>	<mark>0.005 t</mark>
Esters	Ethyl Butanoate	0.287	<mark>0.186 a</mark>	<mark>0.679 a</mark>	<mark>0.530 a</mark>	<mark>7.258 a</mark>
	Ethyl Hexanoate	0.019	<mark>0.080 a</mark>	<mark>0.080 a</mark>	<mark>0.112 a</mark>	0.059 a
	Ethyl 3-hydroxyhexanoate	2.792	<mark>2.414 b</mark>	<mark>2.965 b</mark>	<mark>3.936 a</mark>	<mark>2.765 t</mark>
	Ethyl Acetate	0.812	0.697 ab	<mark>0.503 b</mark>	<mark>0.957 a</mark>	<mark>0.782 a</mark>
Ketones	Acetone	68.129	<mark>87.555 b</mark>	<mark>715.801 a</mark>	101.312 b	134.433 b

*Values followed by the same letter within rows are not significantly different by Duncan's test at the 0.05 level.

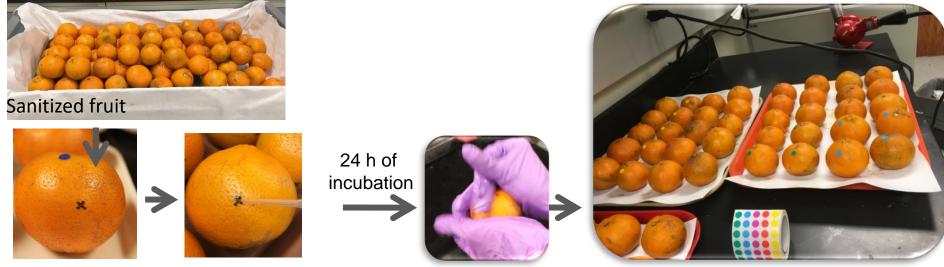
Incorporation of ginger oil into coatings for effect on citrus quality and disease development

- Experiment 2- 'Unique' ortanique tangor ('Valencia' orange/tangerine) quality
 - Repetition of exp.1 with an additional treatment of nanoemulsion containing 0.8% ginger oil stored at 10 °C for 14 d followed by a simulated marketing period (7 d at 20 °C)

Exp.3 - Natural decay and *P. digitatum*-inoculated 'Unique' tangors stored at 20 °C for 31 days and 21 days, respectively, coated with:

1) carnauba nanoemulsion coating; 2) carnauba nanoemulsion coating + 0.8% GEO;

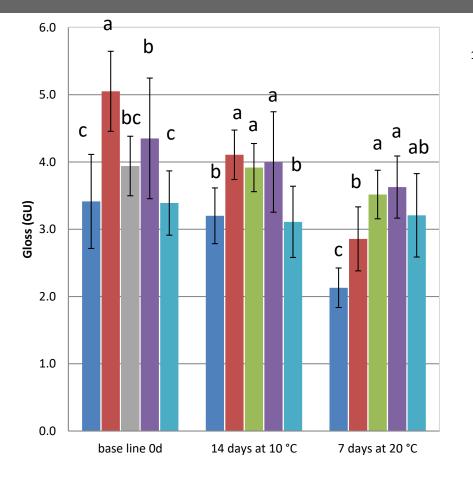
3) carnauba nanoemulsion coating + 0.8% commercial GEO; 4) 0.8% GEO; 5) 0.8% commercial GEO and 6) uncoated/untreated control



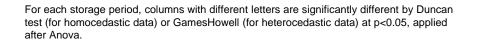
stored at 10 °C/14 d; 20 °C/7 d

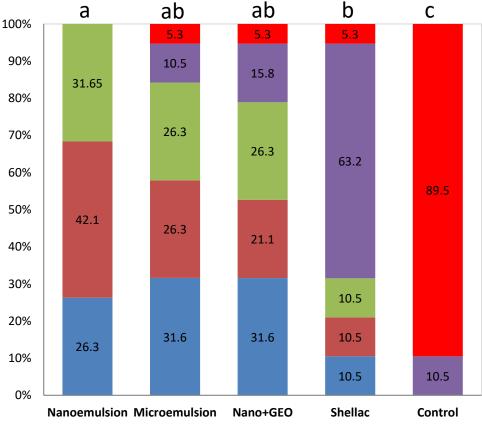
Wound : 1 mmm 7uL spores suspension 10⁶

Exp.2: gloss and sensory shine rank test



■ Control ■ Shellac ■ Nanoemulsion ■ Nano+GEO ■ Microemulsion

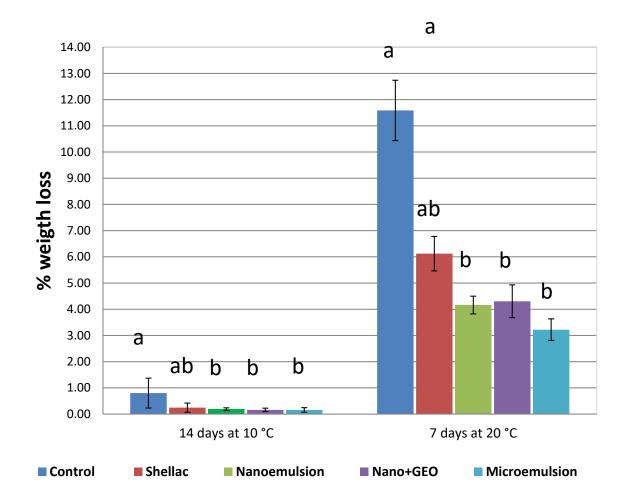




Rank 1 (most shine) Rank 2 Rank3 Rank4 Rank 5 (least shine)

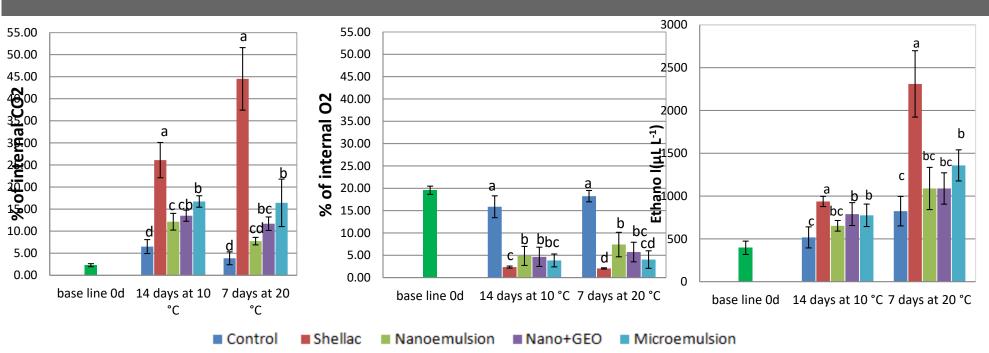
Columns with different letters are significantly different by critical absolute rank sum differences table at p<0.05 (Newell and MacFarlane, 1987),

Exp.2: Weight loss and Sensory firmness



For each storage period, columns with different letters are significantly different by Duncan test (p<0.05), applied after Anova.

Exp.2: Internal CO₂ and O₂ and ethanol



For each storage period, columns with different letters are significantly different by Games Howell test (CO2 and O2 data were hoteroscedastic) or Duncan (homoscedastic data to ethanol) at p<0.05, applied after Anova.

> The lower ethanol level in the nanoemulsion coating +/- GEO, would indicate better flavor compared to shellac or the microemulsion coating, however, uncoated fruit had the lowest ethanol levels and CO_2 along with the highest O_2 (least anaerobic respiration). > GEO did not significantly change the nanoemulsion barrier properties.

Volatiles for Unique Tangors

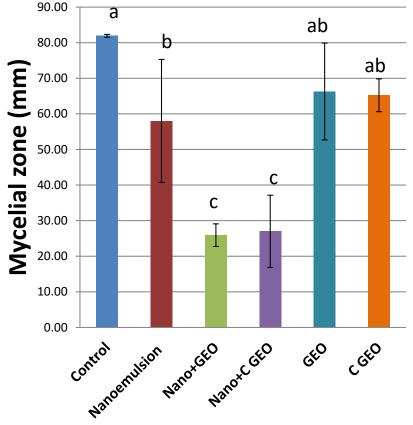
Class	Compound	Base line	Control	Shellac	Nano carnauba	Nano carnauba + GEO	Micro carnauba	Control	Shellac	Nano carnaub a	Nano carnauba + GEO	Micro carnauba
		0 day			14 days at 10	°C		14 days at 10 °C + 7 days at 20 °C				
	Acetaldehyde	0.708	<mark>0.850* a</mark>	<mark>0.514 c</mark>	0.723 b	0.694 b	0.680 b	<mark>0.895 a</mark>	<mark>0.634 b</mark>	<mark>0.850 a</mark>	<mark>0.823 a</mark>	<mark>0.872 a</mark>
A1J-1	Hexanal	0.177	0.293 c	<mark>0.541 a</mark>	0.366 bc	0.435 b	0.405 b	0.237 d	<mark>1.112 a</mark>	0.765 b	0.451 cd	0.698 bc
Aldehydes	Octanal	0.028	0.026 b	<mark>0.031 a</mark>	0.026 b	0.027 b	0.026 b	0.019 d	<mark>0.048 a</mark>	0.028 c	0.021 cd	0.037 b
	Decanal	0.475	<mark>0.436 b</mark>	<mark>0.586 a</mark>	<mark>0.598 a</mark>	0.503 ab	<mark>0.563 a</mark>	0.808 b	<mark>1.921 a</mark>	<mark>0.676 b</mark>	<mark>0.612 b</mark>	0.757 b
	Methanol	4.948	<mark>2.617 a</mark>	<mark>1.971 b</mark>	2.273 ab	2.349 ab	2.253 ab	<mark>2.646 a</mark>	1.666 c	2.362 b	2.571 ab	2.401 b
	Ethanol	431.321	602.892 c	<mark>937.194 a</mark>	652.624 bc	740.511 b	682.339 bc	872.189 d	<mark>2293.789 a</mark>	1190.604 c	1011.583 cd	1407.279 b
	2-MethyPropanol	0.035	0.015 bc	0.025 b	0.020 bc	<mark>0.059 a</mark>	<mark>0.012 c</mark>	0.011 c	<mark>0.165 a</mark>	<mark>0.079 b</mark>	0.014 c	0.029 c
	Hexanol	0.226	0.192 b	<mark>0.278 a</mark>	<mark>0.261 a</mark>	<mark>0.266 a</mark>	0.233 ab	0.212 b	<mark>0.327 a</mark>	0.225 b	0.208 b	<mark>0.349 a</mark>
Alcohols	cis-3-Hexenol	0.573	0.207 c	<mark>0.372 a</mark>	0.283 b	0.267 bc	0.277 bc	0.297 b <mark>c</mark>	<mark>0.984 a</mark>	0.446 b	0.246 c	0.462 b
AICOHOIS	trans-2-Hexenol	0.003	0.005 d	0.012 ab	0.008 c	0.009 bc	<mark>0.012 a</mark>	<mark>0.021 a</mark>	<mark>0.044 b</mark>	<mark>0.026 a</mark>	<mark>0.022 a</mark>	<mark>0.026 a</mark>
	Linalool	0.562	<mark>0.626 a</mark>	<mark>0.907 a</mark>	<mark>0.763 a</mark>	<mark>1.052 a</mark>	<mark>0.772 a</mark>	<mark>0.782 b</mark>	1.126 ab	<mark>1.427 a</mark>	0.745 b	<mark>1.445 a</mark>
	Octanol	0.419	0.218c	0.425 ab	0.333 b <mark>c</mark>	<mark>0.536 a</mark>	0.383 b	0.482 c <mark>d</mark>	<mark>1.211 a</mark>	0.674 bc	<mark>0.423 d</mark>	0.821 b
	Terpinen-4-ol	0.281	<mark>0.240 a</mark>	<mark>0.265 a</mark>	0.288 a	<mark>0.270 a</mark>	<mark>0.254 a</mark>	0.225 c	<mark>0.346 a</mark>	0.259 b <mark>c</mark>	0.241 b <mark>c</mark>	0.311 <mark>a</mark> b
	α-Terpineol	0.154	<mark>0.307 a</mark>	<mark>0.330 a</mark>	<mark>0.342 a</mark>	<mark>0.307 a</mark>	0.209 b	<mark>0.193 a</mark>	<mark>0.243 a</mark>	<mark>0.252 a</mark>	<mark>0.174 a</mark>	<mark>0.201 a</mark>
	α-Pinene	0.107	0.062 c	0.105 ab	0.090 b	<mark>0.114 a</mark>	<mark>0.119 a</mark>	0.120 c <mark>d</mark>	<mark>0.245 a</mark>	0.150 c	<mark>0.108 d</mark>	0.205 b
	Sabinene	0.015	0.009 c	0.016 b	0.013 b	<mark>0.021 a</mark>	0.014 b	0.017 d	<mark>0.048 a</mark>	0.030 bc	0.024 c <mark>d</mark>	0.036 b
Tamanas	Myrcene	0.548	0.313 c	0.544 <mark>a</mark> b	0.475 b	<mark>0.601 a</mark>	<mark>0.626 a</mark>	0.592 c	<mark>1.223 a</mark>	0.764 b <mark>c</mark>	0.627 c	0.901 b
Terpenes	Limonene	11.333	<mark>5.695 c</mark>	10.924 <mark>a</mark> b	9.217 b	11.803 <mark>a</mark> b	<mark>13.097 a</mark>	12.082 c	<mark>32.170 a</mark>	16.750 c	11.882 c	22.600 b
	γ-Terpinene	0.003	0.002 ab	0.002 ab	0.002 b	<mark>0.003 a</mark>	0.002 ab	0.003 c	<mark>0.006 a</mark>	0.004 b <mark>c</mark>	0.002 c	0.005 b
	Valencene	10.162	<mark>7.130 b</mark>	8.930 ab	<mark>9.141 a</mark>	<mark>9.226 a</mark>	8.922 ab	<mark>7.038 b</mark>	9.058 ab	<mark>9.267 a</mark>	8.711 ab	8.136 ab
Esters	Methyl Butanoate	0.038	<mark>0.053 a</mark>	<mark>0.054 a</mark>	<mark>0.015 b</mark>	<mark>0.049 a</mark>	<mark>0.046 a</mark>	<mark>0.052 a</mark>	<mark>0.067 a</mark>	<mark>0.052 a</mark>	<mark>0.055 a</mark>	<mark>0.060 a</mark>
	Ethyl Butanoate	0.047	<mark>0.034 b</mark>	<mark>0.071 a</mark>	<mark>0.029 a</mark>	0.084 b	0.029 b	<mark>0.155 b</mark>	<mark>0.520 b</mark>	<mark>0.224 b</mark>	<mark>0.127 b</mark>	2.419 a
	Ethyl Hexanoate	0.032	0.025 b	0.030 <mark>ab</mark>	0.023 b	<mark>0.034 a</mark>	<mark>0.034 a</mark>	0.044 c	<mark>0.085 a</mark>	0.050 b <mark>c</mark>	0.036 c	0.061 b
	Ethyl 3-hydroxyhexanoate	23.731	<mark>24.494 b</mark>	<mark>32.742 a</mark>	29.672 ab	<mark>36.645 a</mark>	<mark>32.600 a</mark>	24.292 c	<mark>37.992 a</mark>	32.693 b	<mark>37.763 a</mark>	<mark>40.781 a</mark>
	Ethyl Acetate	0.683	<mark>1.250 a</mark>	<mark>1.155 a</mark>	<mark>1.086 a</mark>	<mark>1.096 a</mark>	<mark>1.275a</mark>	<mark>1.294 b</mark>	<mark>2.248 a</mark>	<mark>2.232 a</mark>	<mark>1.498 b</mark>	1.553 b
Ketones	Acetone	149.691	216.931 c	<mark>327.512 a</mark>	236.466 bc	256.123 b	237.134 bc	303.940 d	<mark>721.868 a</mark>	414.981 bc	351.979 c <mark>d</mark>	485.116 b

*For each period of storage, values followed by the same letter within rows are not significantly different by Duncan's test at the 0.05 level.

Exp.3: Coated Petri-dishes

- 10 uL spore suspension 10⁶ was placed on PDA in Petri dishes
- After 24h incubation, 1 mL of each coating from the 3rd expt. was placed and spread on Petri dishes, then incubated for 7 days
- Sterile water were control treatment

For the *in vitro* assay, the nanoemulsion coating improved the antimicrobial activity of the GEO
 The Commercial GEO (C GEO) had similar activity to extracted GEO in this assay

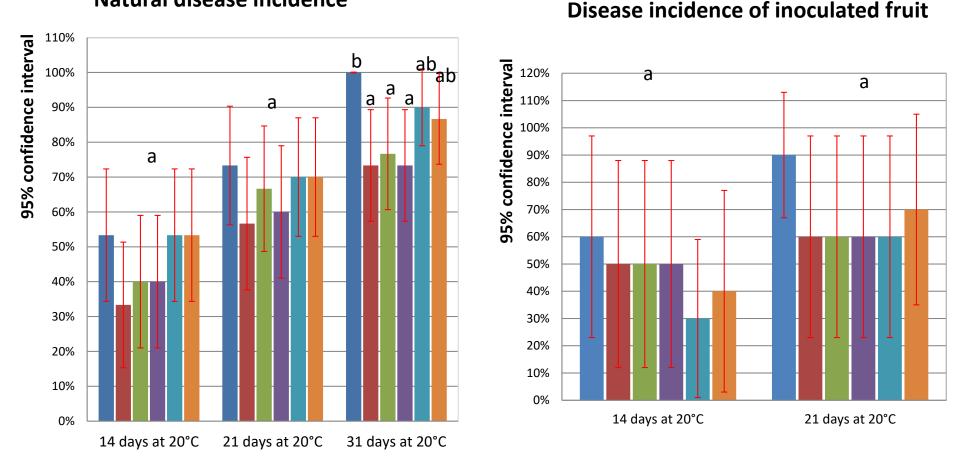


Coated Petri dishes

Columns with different letters are significantly different by Games Howell (p<0.05) applied after Anova.

Exp.3: Natural disease and *P. digitatium* inoculated disease incidence

Natural disease incidence



■ Control ■ Nanoemulsion ■ Nano+GEO ■ Nano+CGEO ■ GEO ■ CGEO

Columns with different letters are significantly different by 95% confidence interval, p<0.05.

Conclusions

> The GEO generally exhibited the most antimicrobial activity among the ginger extracts

>Two times *Peniciulium digitatium* MIC (0.8%) added to nanoemulsion carnauba wax coating was not enough to significantly reduce decay on tangor fruit after 31 d of storage

>The nanoemulsion carnauba coating performed as well as or better than the microemulsion and better than shellac for water loss and formation of off-flavor ethanol indicator

➢ The combination of nanoemulsion + GEO was more effective in coated petri dishes than the nanoemulsion or the GEO alone, but not on fruit, where the nanoemulsion showed antimicrobial ability on its own w/ or w/o GEO and better than GEO alone

Higher concentrations of GEO in nanoemulsion coatings will be tested as GEO shows promising antimicrobial ability for application in edible coatings for fruits

> The coatings did not affect fruit sugar and acid levels, however aroma volatiles from the fruit were analyzed, and showed changes in the aroma profile in addition to ethanol.







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